

.....

Aneuploidy and Cancer: From Correlation to Causation

Peter Duesberg^a, Ruhong Li^a, Alice Fabarius^b, Ruediger Hehlmann^b

^aDepartment of Molecular and Cellular Biology, Donner Laboratory, UC Berkeley, Berkeley, Calif., USA; ^bIII. Medizinische Klinik Mannheim, University of Heidelberg at Mannheim, Mannheim, Germany

Abstract

Conventional genetic theories have failed to explain why cancer (1) is not found in newborns and thus not heritable; (2) develops only years to decades after ‘initiation’ by carcinogens; (3) is caused by non-mutagenic carcinogens; (4) is chromosomally and phenotypically ‘unstable’; (5) carries cancer-specific aneuploidies; (6) evolves polygenic phenotypes; (7) nonselective phenotypes such as multidrug resistance, metastasis or affinity for non-native sites and ‘immortality’ that is not necessary for tumorigenesis; (8) contains no carcinogenic mutations. We propose instead that cancer is a chromosomal disease: Accordingly, carcinogens initiate chromosomal evolutions via unspecific aneuploidies. By unbalancing thousands of genes aneuploidy corrupts teams of proteins that segregate, synthesize and repair chromosomes. Aneuploidy is thus a steady source of karyotypic–phenotypic variations from which, in classical Darwinian terms, selection of cancer-specific aneuploidies encourages the evolution and subsequent malignant ‘progressions’ of cancer cells. The rates of these variations are proportional to the degrees of aneuploidy, and can exceed conventional mutation by 4–7 orders of magnitude. This makes cancer cells new cell ‘species’ with distinct, but unstable karyotypes, rather than mutant cells. The cancer-specific aneuploidies generate complex, malignant phenotypes, through the abnormal dosages of the thousands of genes, just as trisomy 21 generates Down syndrome. Thus cancer is a chromosomal rather than a genetic disease. The chromosomal theory explains (1) nonheritability of cancer, because aneuploidy is not heritable; (2) long ‘neoplastic latencies’ by the low probability of evolving competitive new species; (3) nonselective phenotypes via genes hitchhiking on selective chromosomes, and (4) ‘immortality’, because chromosomal variations neutralize negative mutations and adapt to inhibitory conditions much faster than conventional mutation. Based on this article a similar one, entitled ‘The chromosomal basis of cancer’, has since been published by us in Cellular Oncology 2005;27:293–318.

Copyright © 2006 S. Karger AG, Basel

Despite over 100 years of cancer research, the cause of cancer is still a matter of debate [1–26]. We propose here that the problem of cancer is still

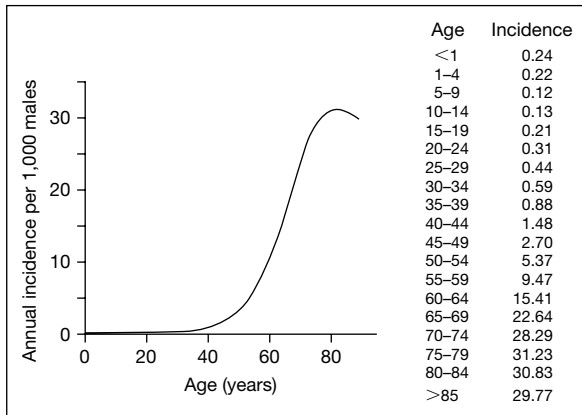


Fig. 1. Age-specific incidence of invasive cancers of males in the United States in 2001. The dominant contributors to the total number of invasive cancers are solid tumors. The growth is approximately exponential until about age 70 and then levels off. Data for the figure, shown in the table at the right, are from the National Program of Cancer Registries at <http://www.cdc.gov/cancer/npcr/index.htm>.

unsolved, because this debate has been monopolized by conventional genetic theories, which hold that cancer is a ‘genetic disease’ [27–35]. But these genetic theories cannot explain any of the following properties of carcinogenesis:

Cancer Is Not Heritable

The best news about cancer is that we and other animals are all born cancer-free and typically acquire cancer, if at all, only at advanced age [34, 36–40]. This bias of cancer for old age is exponential, increasing the cancer risk 300-fold with age, from near-zero rates in newborns and adolescents to rates of 1 in 3 in the last third of a human or animal life span (fig. 1).

In view of the prevailing gene-based cancer theory, however, this age bias is paradoxical. This theory holds that cancer is caused by clonal expansion of one single cell that has accumulated about four to seven complementary mutations during the lifetime of a patient [1, 12, 34, 38, 41, 42]. If this theory is correct, cancer should be common in newborns. For example, a baby, which inherits 3 colon cancer mutations from his mother and 2 from his father, out of the presumably 6 that are thought to cause colon cancer [1, 34], should develop cancer at a very young age from just one more spontaneous mutation in any one of the billions of its colon cells. Indeed, many hypothetical cancer-causing mutations, including those thought to cause colon cancer, are heritable in transgenic mice (Appendix) and also in humans. According to Vogelstein and Kinzler [43], “one

of the cardinal principles of modern cancer research is that the same genes cause both inherited and sporadic (noninherited) forms of the same tumors”.

But there is no colon cancer in newborns (fig. 1). Thus, cancer is somatically generated and not a heritable disease.

Long Neoplastic Latencies

Experimental or accidental carcinogenesis, and the age bias, demonstrate that cancer is a late product of a gradual evolution of somatic cells that may be ‘initiated’ either by carcinogens or spontaneously [1, 10, 38, 40, 44, 45]. Once initiated, this evolution is autonomous but very slow, generating cancer cells only after lengthy and uneventful ‘neoplastic latencies’ [40, 45]. These latencies last many months to years in carcinogen-treated rodents and decades in accidentally exposed humans [40, 45–48]. For example, (1) the solid cancers, which developed in human survivors only 20 years after the explosion of atomic bombs in Japan in 1945 [38]; (2) the breast cancers, which developed only 15 years after treatments of tuberculosis with X-rays in the US in the 1950s [49], and (3) the lung cancers, which developed in workers of a mustard gas factory only 30 years after it was closed in Japan in 1945 [50]. The exponential increase of the spontaneous cancer risk of humans with age even implies neoplastic latencies of up to 50 years from a near zero-risk at birth to a one in three risk in the last three decades of a human lifespan of about 80 years (fig. 1). The primary cancer cells that appear after these lengthy pre-neoplastic evolutions continue to progress independently within individuals tumors to form evermore ‘polymorphic’ [51] and malignant cancers with evermore exotic karyotypes and phenotypes [45].

These long latencies of carcinogenesis, however, are incompatible with the immediate effects of conventional mutation [2, 31, 35, 52]. It is for this reason that Cairns wrote in *Cancer: Science and Society*: ‘The conspicuous feature of most forms of carcinogenesis is the long period that elapses between initial application of the carcinogen and the time the first cancers appear. Clearly, we cannot claim to know what turns a cell into a cancer cell until we understand why the time course of carcinogenesis is almost always so extraordinarily long’ [38].

Non-Mutagenic Carcinogens Cause Cancer

Both mutagenic and non-mutagenic carcinogens cause cancer. Examples of non-mutagenic carcinogens are asbestos, tar, mineral oils, naphthalene, polycyclic aromatic hydrocarbons, butter yellow, urethane, dioxin, hormones, metal ions such as Ni, Cd, Cr, As, as well as spindle blockers such as vincristine and colcemid, extranuclear radiation and solid plastic or metal implants (Appendix). Conventional genetic theories, however, fail to explain carcinogenesis by non-mutagenic carcinogens.

Karyotype-Phenotype Variations at Rates that Are Orders Higher than Mutation

During the neoplastic phase of carcinogenesis, cancer cells gain or lose chromosomes or segments of chromosomes (fig. 2) and change phenotypes at rates that far exceed those at which genotypes and phenotypes are changed by conventional mutation [53–55]. For example, highly aneuploid cancer cells become drug resistant at rates of up to 10^{-3} per cell generation [53, 54, 56–58] or become metastatic at ‘high rates’ [59, 60]. As a result of this inherent chromosomal instability most cancers are enormously heterogeneous populations of nonclonal and partially clonal, or sub-clonal cells [13, 61]. Thus, cells from the same cancer differ from each other in ‘bewildering’ phenotypic and chromosomal variations [62] and in mutations – even though most cancers are derived from a common, primary cancer cell and thus have clonal origins [38, 45, 51, 56, 61, 63–67].

By contrast, the karyotypes of normal cells are stable despite mutational or developmental phenotype variations [31, 34, 52, 68]. And phenotypic variation of normal cells by conventional gene mutation cells is limited to 10^{-7} per cell generation for dominant genes and to 10^{-14} for pairs of recessive genes in all species [6, 47, 52, 57, 68, 69]. Even the mutation rates of most cancers are not higher than those of normal cells [6, 19, 20, 47, 66, 70–75]. Thus, phenotypic variation in cancer cells can be four to eleven orders faster than conventional mutation.

Cancer-Specific Aneuploidies

Despite the karyotypic instability and heterogeneity of cancer cells partially specific or nonrandom aneuploidies have been found in cancers since in the late 1960s [61, 62, 76–87]. Since the 1990s, many more nonrandom aneuploidies have been detected in cancers by the use of comparative genomic hybridization, rather than by identifying specific aneusomies cytogenetically [61, 88–96]. The term aneusomy is used for a specific, aneuploid chromosome. Specific aneuploidies have even been linked with specific stages of carcinogenesis and with specific phenotypes of cancers such as: (1) Distinct stages of neoplastic transformation in human [62, 89, 95–99] and in animal carcinogenesis [84]; (2) invasiveness [97, 98, 100]; (3) metastasis [101–106]; (4) drug-resistance [53, 69, 107]; (5) transplantability to foreign hosts [108]; (6) distinct cellular morphologies [109]; (7) abnormal metabolism [62, 110], and (8) cancer-specific receptors for viruses [62, 109].

Cancer-specific, nonrandom aneuploidies, however, are inconsistent with the conventional mutational theories of cancer. In fact they are a direct challenge of the mutation theory, because specific aneusomies have the potential to generate cancer-specific functions (Appendix). The Down syndrome-specific functions of trisomy 21 are a confirmed model [111–114].

Karyotypes of clonal cultures of the near-diploid human colon cancer line HCT 116 and the hyper-diploid human colon cancer line SW480																					
HCT 116, mn=45			SW480, mn=57																		
Chrom.	Metaphases		Chrom.	Metaphases																	
	1 to 29	30		1 to 6	10	15	19														
1	2	2	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
3	2	2	3	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
4	2	2	4	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
5	2	2	5	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
6	2	2	6	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
7	2	2	7	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
8	2	2	8	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
9	2	2	9	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0
10	1	1	10	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
11	2	2	11	3	3	3	3	3	3	3	3	3	3	2	3	3	3	3	3	3	3
12	2	2	12	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1
13	2	2	13	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3
14	2	2	14	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
15	2	2	15	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
16	1	1	16	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
17	2	2	17	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3	3
18	1	1	18	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
19	2	2	19	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
20	2	2	20	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
21	2	2	21	3	3	3	3	3	3	3	3	3	3	2	3	3	3	3	3	3	1
22	2	2	22	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2	2
X	1	1	X	2	2	2	2	2	2	2	2	2	2	1	2	2	2	2	2	2	2
Y	0	0	M1 2/12	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
M1 10 ⁺	1	1	M2 3/12/10	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
M2 8/16	1	1	M3 9/1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
M3 17/18	1	1	M4 Ω9/1	1	1	1	0	0	0	0	0	0	0	0	1	1	1	1	1	1	1
M4 12 ⁻	0	1	M5 3 ⁺	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	1	1	0
			M6 8/9	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M7 7/14	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M8 5/20/7	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	0
			M9 5/20	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M10 Ω5/20	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M11 3 ⁻	1	1	1	1	1	1	1	0	1	1	1	1	1	1	1	1	1	2
			M12 12 ⁻	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	2
			M13 19/8/19/5	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M14 19/8	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1	1
			M15 15/18	2	1	1	2	1	2	2	2	1	2	2	2	2	2	2	2	2	2
			M16 16/14/13	0	0	0	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0
			M17 9/5	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0	0
			M18 2/8	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
			M19 9/1/11	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0
			M20 12/1	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0	0
			M21 21/11	0	0	0	0	0	0	0	1	0	0	0	0	0	0	0	0	0	0

Fig. 2. Karyotypes of clonal cultures of human colon cancer and Chinese hamster cell lines. **a** Karyotypes of clonal cultures of the near-diploid human colon cancer cell line HCT 116 (modal chromosome number = 45) and of the hyper-diploid human colon cancer cell line SW480 (modal chromosome number = 57). The karyotype of only 1 out of 30 cells of the clonal culture of the near-diploid HCT 116 line was non-clonal, containing an extra, partially deleted chromosome 12, termed marker M4 12⁻ (**bold italic** number). By contrast, 13 (**bold italic** numbers) out of 19 cells of the clonal culture of the hyper-diploid SW480 line had nonclonal karyotypes. All 13 nonclonal karyotypes differed from the modal karyotype of this line in the numbers of one or more chromosomes. Four of these 13 nonclonal cells also contained new structurally altered chromosomes, labeled M16 to 21 (**bold italic** numbers). Chromosomal constituents of the marker (hybrid) chromosomes are indicated following their

Karyotypes of clonal cultures of the near-diploid, hyper-diploid and near-triploid Chinese hamster cells																	
Clone	Meta	Chr No.	Normal chromosomes										Altered chromosomes				
			1	2	3	4	5	6	7	8	9	10	X	Y	Partially clonal	Non-clonal	
B69-1 mn = 23	1 to 17	23												1	ac1-2		
	18	24												1	ac1-2	ac101	
	19	24												1	ac1-2	ac102	
	20	48	4	4	4	4	4	4	4	4	4	4	4	2	2	2	ac1[2]-2[2]
D1 mn = 29	1	30		4		3	4	4							ac1		
	2	29		4		3	3	4							ac2		
	3	28		4	3		3	3							ac1		
	4	29		4		3	3	4							ac2		
	5	30		3		3	4	4							ac1-2		
	6	30		4	3		4	4							ac1		
	7	29		4		3	4	4			0				ac1		
	8	29		4			4	4							ac1		
	9	28		4			3	3							ac1-2		
	10	29		4			4	4								ac101	
	11	30		4			3	4	4						ac1		
	12	33		4			3	4	4						ac2	ac102-104	
	13	30		4			4	4			3				ac1		
	14	27		4			3	3			3						
	15	32		4		3	3	4	4	3						ac105	
	16	30		4			3	4	4						ac1		
17	30		4				4	4						ac1	ac106		
18	29		4			3	4	4									
19	32		4	4			4	4							ac102, 107		
20	29		4		1	3	4	4						ac1			
B2 mn = 35	1	33						3	3	3					ac1, 5, 7, 12, 21-22	ac201-202	
	2	34							3						ac1-2, 4-5, 7, 21, 23	ac203-206	
	3	34			1										ac1-2, 4-5, 11, 21-22	ac207-212	
	4	34													ac1-2, 4-5, 7, 12, 22, 24	ac213-216	
	5	33					3								ac2, 4, 5, 7, 22, 24	ac217-220	
	6	38					3	1							ac1, 4-5, 7, 21-22	ac221-230	
	7	34					3								ac1, 4, 7, 11-12, 22	ac231-235	
	8	34				3	3								ac1-4, 7, 12, 21, 24	ac236-237	
	9	36					3				3				ac2, 5, 7-8, 11-12, 22	ac238-242	
	10	33													ac1, 3, 11-12, 22, 24	ac243-247	
	11	32	1	1		1		1							ac1-2, 4-5, 7, 11, 21	ac248-254	
	12	36					3	3							ac1-2, 4-5, 11, 21, 24	ac255-259	
	13	37					3		3	3					ac1-5, 7[2]-8, 22	ac260-262	
	14	38					3								ac1-2, 4-5, 7-8, 12, 21	ac263-269	
	15	32					3	3							ac1, 4-5, 7, 22	ac270-272	
	16	30	1	1											ac1, 5, 7, 21	ac273-278	
17	32				3									ac1, 4-7, 12	ac279-281		
18	34													ac2-5, 11, 21	ac282-287		
19	36					3			3	4				ac1-2, 4, 12, 21-22	ac288-291		
20	34													ac1-2, 4-5, 12, 21-22	ac292-296		
21 to 26	~66														too complex to analyze		

designation, e.g. M1 2/12 for a hybrid of chromosomes 2 and 12. **b** Karyotypes of clonal cultures of the near-diploid, hyper-diploid and near-triploid Chinese hamster cell lines B69-1 (modal chromosome number = 21), D1 (modal chromosome number = 29) and B2 (modal chromosome number = 35). No numbers signal normal chromosome numbers. It can be seen that only 3 of 20 cells of the near-diploid line B69-1 had nonclonal karyotypes. Each of these included one new structurally altered chromosome, termed ac101 and ac102. One of these three nonclonal karyotypes also had undergone tetraploidization. By contrast, there were no two identical cells in the clonal cultures derived from the hyper-diploid and near-triploid Chinese hamster cells. Nevertheless, the degrees of both numerical and structural variations were much higher in near-triploid than in hyper-diploid Chinese hamster cells.

Cancers Have Complex Phenotypes

The complexity of most cancer-specific phenotypes far exceeds that of phenotypes generated by conventional mutation. For example, the kind of drug-resistance that is acquired by most cancer cells exposed to a single cytotoxic drug is more complex than just resistance against the drug used to induce it. It protects not only against the toxicity of the challenging drug, but also against many other chemically unrelated drugs [56, 58, 115]. Therefore, this phenotype has been termed ‘multidrug resistance’. Thus, drug resistance must be polygenic. The same is likely to be true for the other cancer-specific phenotypes such as grossly altered metabolism, invasiveness, metastasis, and immortality [40, 45], because all of these phenotypes correlate with altered expressions of thousands of genes [34, 87, 116–118] and with highly abnormal concentrations of thousands of normal proteins [16, 40, 51, 119]. Moreover, in highly aneuploid cancer cells the number of centrosomes is increased up to 5-fold – from a normal of two to around ten – and at the same time their structures are often altered [120–123].

The high genetic complexities of most cancer-specific phenotypes, however, are incompatible with accumulations of large numbers of gene mutations generated at conventional rates during the limited live spans of humans and animals. Indeed, it is virtually impossible that the up to 5-fold increased numbers of centrosomes that are observed in highly aneuploid cancer cells [17, 120, 121, 124], would be the result of mutations that increase the numbers of the 350 different proteins that make up centrosomes [125].

Nonselective Phenotypes of Cancer Cells

Cancer-specific phenotypes can be divided into two classes: Those, which are selective, because they advance carcinogenesis by conferring growth advantages to cancer cells such as invasiveness, grossly altered metabolism and high adaptability via high genomic variability [40, 45], and those, which are not selective for growth [73, 126]. The nonselective, cancer-specific phenotypes include metastasis, drug resistance and immortality. Metastasis is the ability to grow at a site away from the primary tumor. Therefore, it is not selective at the site of its origin [126]. Likewise, drug resistance is not a selective advantage for natural carcinogenesis in the absence of chemotherapy. Yet, a high percentage of cancers is a priori or intrinsically drug-resistant [127, 128]. Moreover, the majority of the drug resistances associated with multidrug resistance offer no selective advantages against the drug that induced it. Even immortality is not a selective advantage for carcinogenesis, because many types of human cells can grow over 50 generations according to the Hayflick limit [129], and thus many more generations than are necessary to generate a lethal cancer. Consider that 50 cell generations produce from one single cell a cellular mass equivalent of 10 humans with 10^{14} cells each [10]. Nonselective

phenotypes, however, are entirely inconsistent with conventional gene mutation-selection mechanisms.

No Carcinogenic Genes in Cancer

Numerous gene mutations have been found in cancer cells since the 1980s [1, 29, 42, 130–133], and the prevailing genetic theories of cancer postulate that these mutations are carcinogenic [29, 30, 33, 34, 42].

But none of the mutations found in cancers are cancer-specific [1, 134], and in cases where this information is available many, perhaps most, mutations are nonclonal [8, 134, 135] and are not detectably expressed in human cancer cells in vivo [8, 116, 136, 137]. Despite enormous efforts in the last 25 years, no mutant gene and no combination of mutant genes from cancer cells has been found that converts diploid human or animal cells into cancer cells [4, 5, 12, 13, 24, 73, 138]. Moreover, mouse strains with artificially implanted, hypothetical cancer genes, or with artificially deleted tumor suppressor genes have survived many generations in laboratories with either the same or slightly higher cancer risks than other laboratory mice (Appendix) [8, 24, 73].

In view of this, Vogelstein and Kinzler [1] closed a very influential review of the mutation theory in 1993 as follows: ‘The genetics of cancer forces us to re-examine our simple notions of causality, such as those embodied in Koch’s postulates: How does one come to grips with words like “necessary” and “sufficient” when more than one mutation is required to produce a phenotype and when that phenotype can be produced by different mutant genes in various combinations?’ These and other inconsistencies between carcinogenesis and established genetic theories are the reasons why it is still debated, whether mutations or aneuploidies or epigenetic alterations cause cancer [1, 3–8, 10–14, 16–22, 24–26, 42].

A New, Chromosomal Evolution Theory of Carcinogenesis

In an effort to resolve the many discrepancies between carcinogenesis and conventional genetic theories listed above, we present here a new, chromosomal evolution theory of carcinogenesis. Our theory is based on: (1) the ubiquity of aneuploidy in cancer [61, 62, 65, 78, 139]; (2) our own data that aneuploidy changes the numbers and structures of chromosomes and phenotypes automatically much faster than and independent of mutation [53–55, 137, 140]; (3) an earlier chromosomal theory of cancer proposed by Boveri and von Hansemann over 100 years ago [141–143]. This theory, however, was abandoned in the 1950s and 1960s in favor of mutation, because instead of the expected cancer-specific aneuploidy, karyotypic heterogeneity was found in most cancers by the methods developed at that time [62, 144, 145]. Ever since, ‘aneuploidy and other forms of

chromosomal abnormality' of cancer cells [56] are generally interpreted as 'secondary' events [24, 56, 61, 62, 146] – secondary to presumably primary gene mutations [15, 32, 64, 75, 147–153]; (4) cancer-specific aneuploidies discovered since the late 1960s by many laboratories including ours, particularly by comparative genomic hybridizations [84]. These discoveries, however, are not appreciated as chromosomal causes of cancer because of the prevailing genetic theories.

According to our new chromosomal evolution theory, carcinogenesis is the result of the following chain of events: (1) carcinogens and spontaneous mitotic errors induce unspecific aneuploidies; (2) aneuploidy corrupts teams of proteins that segregate, synthesize and repair chromosomes. Aneuploidy is thus a steady source of karyotypic-phenotypic variations from which, in classical Darwinian terms, selection of cancer-specific aneuploidies encourages the evolution and spontaneous 'progressions' of the malignant phenotypes of neoplastic cells. The rates of these variations are proportional to the degrees of aneuploidy; (3) this chromosomal evolution makes cancer cells new, inherently unstable cell 'species' with distinct, but unstable karyotypes, rather than mutant cells. Owing to this inherent chromosomal instability, cancers are uncertain combinations of random and of relatively specific or 'nonrandom' aneuploidies; (4) the cancer-specific aneuploidies generate complex, malignant phenotypes via abnormal dosages of thousands of genes. Down syndrome is a model for how aneuploidy generates complex, abnormal phenotypes, and (5) thus cancer is a chromosomal rather than a genetic disease.

Below, we offer a brief explanation of how aneuploidy generates new phenotypes, independent of mutation. According to this mechanism variations of chromosomes have the same effects on the phenotypes of cells as variations of the assembly lines of a car factory on the phenotypes of an automobile. If changes are made that do not alter the balance of components, e.g. moving the engine from the front to the rear, new, competitive car models are generated. Indeed, motor companies change their assembly lines to create a new car model. Likewise, phylogenesis generates new species by changing the numbers and structures of the chromosomes of existing species [154].

If unbalanced, i.e. aneuploid, changes are made, abnormal and defective products must be expected. The human trisomy 21, which causes Down syndrome, is a classic non-neoplastic example [113, 114]. Although trisomy 21 is only a tiny aneuploidy compared to that of most cancers, it generates 71 Down-specific phenotypes [111, 112]. Likewise, experimentally induced, congenital aneuploidies generate numerous abnormal phenotypes in drosophila, plants and mice, independent of gene mutation [155–157]. Thus, the complex aneuploidies of cancer cells can be expected to generate numerous new phenotypes.

By contrast, the power of changing the phenotypes of the cell by gene mutation is comparable to employing a few defective or overactive workers on

the assembly lines of a car factory. Neither of these variables will generate a new car model, except possibly to produce either a defective car or no car at all, if an assembly line comes to a stop [158]. For example, none of the 1.42 million point mutations that distinguish any two humans [159] have generated a new human species, nor have they even been sufficient to cause cancer in newborns.

Instead of being controlled by hypothetical oncogenes or tumor suppressor genes, alias ‘gate keepers and caretakers’ [75, 160], or being de-controlled by the corresponding mutations, most phenotypes of normal and cancer cells are controlled ‘democratically’ by hundreds of kinetically linked proteins [161]. Such cooperative assembly lines of gene products are buffered against mutations of single genes by the assembly line principle [161, 162]. According to this principle, unchanging supplies and demands of numerous unmutated genes from upstream and downstream of biochemical assembly lines buffer mutations in two ways. They automatically raise substrate concentrations upstream of slow-working, mutationally compromised genes and restrict by normal supplies of substrates mutationally activated genes [161, 163]. This is indeed the principle that buffers cells of all multicellular organisms against all but knock out mutations that occur during their long lifetimes.

Thus aneuploidization, upsetting the balance of thousands of normal genes, rather than mutation of a few genes, is necessary to generate the complex and dominant phenotypes of cancer cells.

In sum, the chromosomal evolution theory provides a coherent explanation of carcinogenesis that is independent of mutation, and that can explain each of the many idiosyncratic features of carcinogenesis that are paradoxical in view of the mutation theory. However, the chromosomal theory remains challenged by competing claims of the prevailing genetic theories of cancer. In the following we take up this challenge.

Testing Specific Predictions of the Chromosomal Theory against Competing Claims by Genetic Theories of Cancer

According to the prevailing genetic theories of cancer, ‘carcinogens are mutagens’ [164] initiating carcinogenesis by mutation, and ‘initiated’ cells then evolve into cancer cells via poorly defined sets of four to seven complementary mutations [1, 29, 34, 35, 38, 41, 42, 52, 134, 165]. Since these claims of the prevailing genetic theories of cancer have monopolized cancer research in the last decades, we have tested the most distinctive predictions of the chromosomal evolution theory: (1) carcinogens initiate carcinogenesis by aneuploidisation; (2) aneuploidy is inherently variable and thus sufficient to catalyze the evolution of cancer-specific chromosome patterns, and (3) carcinogenesis is independent of somatic mutation.

Carcinogens Function as Aneuploidogens

This prediction has been confirmed previously by others [4, 10, 44, 67, 70, 73, 158, 166–168] including Boveri, who first demonstrated that X-rays, several chemicals, heat and physical stress generate aneuploidy, but failed to observe cancer in experimental animals [142, 143]. However, since these studies did not establish pre-neoplastic aneuploidy as the cause of carcinogenesis [6, 7, 24, 25], we have recently retested the question whether carcinogens cause aneuploidy experimentally, using mutagenic [84] and nonmutagenic carcinogens [169, 170], and by reviewing the literature [4, 10, 25, 73, 158]. These tests have shown that mutagenic carcinogens generate aneuploidy either by breaking and rearranging chromosomal DNA or by chromosome nondysjunction owing to alterations of the spindle apparatus. By contrast, nonmutagenic carcinogens would induce aneuploidy primarily via de-polymerization of the proteins of the spindle apparatus or even via physical interference with mitosis as by asbestos [80]. Polycyclic aromatic hydrocarbons and vincristine are examples of carcinogens that cause aneuploidy by depolymerizing protein polymers of the spindle apparatus [70, 158].

Moreover, carcinogens, particularly radiations and mutagenic chemical carcinogens, induce aneuploidy without delay, and thus long before cancer [170–174], as postulated by the chromosomal theory. Most importantly, our own studies have shown that among the many effects that carcinogens have on cells [40], aneuploidy is the one that consistently segregates with subsequent carcinogenesis [84, 170].

A series of recent studies, aiming at the definition of mutations that might ‘initiate’ carcinogenesis, have instead all pointed to chromosomal initiation [67, 73, 174]. Based on the dosage of a carcinogen delivered to cell cultures, the percentages of ‘initiated’ cells were found to be >1,000-fold larger than expected for the target gene [73]. Markers identified for the initiation of carcinogenesis were either aneuploidy or chromosomal destabilization or immortalization or ‘delayed reproductive death’ [67] or transformation of cells in vitro [73]. Since an average human chromosome contains about 1,500 genes – 35,000 genes divided by 23 chromosomes [154] – it follows that the chromosome is the target for the initiation of carcinogenesis [73]. We conclude that carcinogens function as aneuploidogens as postulated by the chromosomal theory.

Aneuploidy Is Inherently Variable and Thus Sufficient to Catalyze the Evolution of Cancer-Specific Chromosome Patterns

We have tested this critical prediction of the chromosomal evolution theory, by measuring the rates at which karyotypes of cancer cells vary spontaneously per cell generation. For this purpose clonal cultures of cancer cells with different degrees of aneuploidy were prepared and the fraction of nonclonal karyotypes in these cultures was determined. The rates of karyotype alteration

per cell generation are then calculated by dividing these fractions by the number of generations of the clonal culture.

Using this method we found karyotypic variation at rates of near 10^{-2} per generation in the hyper-diploid – modal chromosome number = 57 – human colon cancer cell line SW480 [53]. This rate was calculated from the data shown in figure 2a as follows: 6 of the 19 karyotypes were identical and are thus considered the ‘stemline’ [62] or modal karyotype of this line. But, 13 of 19 ‘clonal’ SW480 cells had non-clonal karyotypes, differing from the predominant ‘stemline’ in numerical and structural aneusomies, which are identified by bold italic numbers in figure 2a. Since the clone was about 23 generations old by the time it was analyzed, having grown from a single cell to about 10^7 , the average rate of karyotype variation per cell per generation is about 3% (13:19:23). Indeed, this is a minimal estimate, because many random chromosomal variations are not viable. A comparison of the karyotypes of an SW480 cell with a normal human foreskin cell is shown in figure 3. The karyotypes were prepared from metaphase chromosomes hybridized in situ with color-coded chromosome-specific DNA probes, as described by us recently [53].

Even higher rates of over 1 chromosomal variation per cell generation were observed in the hyper-diploid and near-triploid Chinese hamster cell lines D1 (modal chromosome number = 29) and B2 (modal chromosome number = 35) [55, 140] (fig. 2b). The normal chromosome number of the Chinese hamster is 22. Not even two of these highly aneuploid Chinese hamster cells were the same [55]. This means that the rates of karyotype variations per cell generation were at least 4% (100%: 23), but probably higher, because most random variations are likely to be lost as fast as they are generated. However, in the case of the near-triploid B2 line the rates of structural chromosomal rearrangements were at least 100% per generation, because each metaphase contained several unique structural chromosome alterations, numbered ac201-ac296 in figure 2b.

As predicted by the chromosomal theory, much lower rates of karyotype variations were observed at low degrees of aneuploidy, namely in the near-diploid human colon cancer cell line HCT 116 (modal chromosome number = 45) and in the near-diploid Chinese hamster line B69–1 (modal chromosome number = 23) [55, 140]. Only 1 of 30 clonal HCT 116 cells contained a new, structurally altered chromosome, again identified by a bold italic number in figure 2a, which corresponds to a rate of only 0.15% karyotypic variations per cell generation. Not even one purely numerical variation was detected in 30 metaphases. Likewise only 3 of 20 clonal B69–1 cells had nonclonal karyotypes (fig. 2b), which corresponds to a rate of 0.65% karyotypic variations per cell generation.

It follows that the degrees of both numerical and structural chromosomal instability of human and Chinese hamster cells are proportional to the degrees

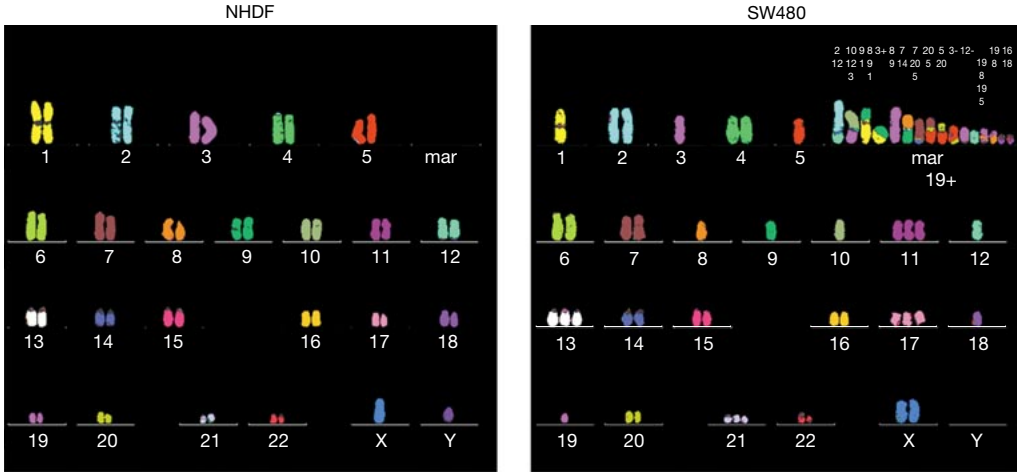


Fig. 3. Metaphase chromosomes of a normal human foreskin cell and of a cell from the human colon cancer cell line SW480. Cytogenetically intact chromosomes are identified by numbers. The group labelled 'mar' (for marker chromosome) shows structurally abnormal chromosomes, which are either rearranged intra-chromosomally or inter-chromosomally to form various hybrid chromosomes. The numbers above these marker chromosomes identify the chromosomal origins of hybrid chromosomes in their relative order or the basis of intra-chromosomal alterations, e.g. 3+ for an amplification of chromosome 3. A comparison of the two karyotypes shows that the cancer cells differ from the normal cell in numerous numerical and structural chromosomal alterations or aneusomies. See online version for color.

of aneuploidy, as postulated by the chromosomal theory. Others have recently described very similar correlations between chromosomal instability and degrees of aneuploidy in human cancer cells including some of those used by us [175–177].

However, the fact that chromosomes are destabilized in proportion to the degree of aneuploidy could also be explained by a series of independent mutations. But, this mutation argument is unlikely, because it is very unlikely that two inherently different kinds of mutations, those that alter the structures and those that alter numbers of chromosomes, would both be equally proportional to the degrees of aneuploidy in all cancers, considering that specific mutations are very rare, even in cancer cells (Appendix). In other words, this argument predicts some cancers with high numerical and no or low structural instability, and others with the opposite distribution, but so far no such cancers have been described.

In sum, the conclusion can be drawn that the inherent variability of aneuploidy is the cause of the chromosomal and phenotypic instabilities of cancer cells and the resulting cellular heterogeneities of cancer, as predicted by the

chromosomal theory. This aneuploidy-specific, chromosomal uncertainty principle had become the nemesis of the Boveri-von Hansemann theory in the 1950s and 1960s.

Carcinogenesis Independent of Somatic Mutation

Cancer coincides with aneuploidy as well as with mutations [6, 7, 10, 13, 24]. In the words of a recent review in *Science*, ‘Cancer cells are chock-full of mutations and chromosomal abnormalities’ [6]. Therefore, it can be argued that spontaneous and carcinogen-induced aneuploidization is sufficient for the initiation and autocatalytic evolution of carcinogenesis, as we did here. But, it could also be argued that the initial aneuploidization and its subsequent evolution depend on somatic mutations, as others have done recently [13, 14, 26, 150–153, 178].

However, the following 4 arguments indicate that carcinogenesis (of normal cells in normal organisms) is independent of somatic mutation [25]. In fact, cancer cells, via their specific aneuploidy, are even protected against the negative effects of mutation: (1) Initiation of carcinogenesis by aneuploidy, generated by mutagenic carcinogens fragmenting or eliminating chromosomes, is about 35,000 times more likely than by aneuploidy, generated by mutation of a specific mammalian ‘aneuploidy-gene’ [6]. This is because mammals contain about 35,000 genes, and thus only 1 in 35,000 specific mutations would generate an ‘aneuploidy gene’ [25, 154], but any mutation leading to a chromosome break or rearrangement generates aneuploidy. Using nonmutagenic carcinogens to generate initiating aneuploidy via the spindle apparatus is in fact infinitely more efficient than via the nontarget gene. Thus, initiation of carcinogenesis is independent of somatic mutation. (2) Generating the complex, cancer-specific phenotypes by chromosomal variation is about 1,500 times more efficient than by mutation. Indeed, it would be almost impossible to generate the complex, polygenic phenotypes of cancer cells in a lifetime of a cancer patient by mutating many genes, considering the complexity of cancer-specific phenotypes and the low rates of spontaneous mutation in normal and most cancer cells (Appendix). By contrast, chromosomal variation is a mechanism that automatically alters the dosages and expressions of thousands of genes. Therefore, aneuploidization is infinitely more efficient in generating the complex phenotypes of cancer cells than mutation. Thus, carcinogenesis is independent of somatic mutation in generating complex, cancer-specific phenotypes. (3) The high rates of cancer-specific karyotype-phenotype variations are irreconcilable with the low rates of conventional mutation. New, cancer-specific phenotypes appear or old ones disappear in highly aneuploid cancer cells at rates of up to 10^{-3} per cell generation, which is four to eleven orders faster than conventional gene mutation (Appendix). Thus phenotype variation in cancer cells is independent

of mutation. (4) The relevance of somatic mutations for carcinogenesis is uncertain. Cancer-specific aneuploidy can generate gene mutations by the same mechanism that varies the structures of chromosomes. In addition, aneuploidy renders DNA synthesis error-prone by unbalancing nucleotide pools [179]. Thus, the simplest explanation of the many mutations of cancer cells would be that these mutations are consequences of aneuploidy and thus not necessary for carcinogenesis. This hypothesis explains why the mutations found in cancer cells are frequently nonclonal in cancers [8, 135], and why they do not transform normal cells to cancer cells and do not breach the livelihood of transgenic mice (Appendix). Indeed, cancer cells are immortal, because frequent, aneuploidy-catalyzed karyotypic variations neutralize all potentially negative mutations at much higher rates than they can be generated.

We conclude that carcinogenesis is independent of somatic mutation, because aneuploidy is much more likely to be generated and varied at the chromosomal level than by mutation. In response to this it has been argued that cancers associated with heritable cancer-disposition syndromes prove that carcinogenesis is dependent on mutation. Examples are the retinoblastoma, xeroderma, Bloom syndrome, and mosaic variegated aneuploidy syndromes [32, 34, 180, 181]. However, these heritable – rather than somatic – mutations are not direct causes of cancer. Instead they initiate carcinogenesis by aneuploidization at much higher rates than it would occur in normal cells by spontaneous or carcinogen-induced aneuploidization [181–183]. According to the chromosomal theory these mutations are genetic equivalents of carcinogens that induce aneuploidy at high rates. This view is supported by the presence of aneuploidy in such patients prior to carcinogenesis, as for example in mosaic variegated aneuploidy patients [183, 184], Bloom patients [182] and xeroderma patients [185], and by the presence of aneuploidy in the cancers of patients with retinoblastoma [186–189], mosaic variegated aneuploidy [183, 184], xeroderma [185, 190] and Bloom patients [182].

We conclude that the abnormally high rates of carcinogenesis in heritable cancer disposition syndromes are dependent on abnormally high rates of aneuploidizations that are generated by these heritable genes. Thus carcinogenesis encouraged by certain heritable mutations confirms and extends the chromosomal theory of carcinogenesis, but does not show that carcinogenesis in normal cells depends on conventional mutation.

Explanatory Value of the Chromosomal Theory of Cancer

In table 1, we have summarized how the chromosomal cancer theory explains each of the idiosyncratic features of carcinogenesis that are paradoxical

Table 1. Features of carcinogenesis

Genetic paradox	Chromosomal solution
1 Cancer not heritable	aneuploidy is not heritable
2 Long neoplastic latencies	autocatalyzed evolution of cancer-specific aneusomies
3 Non-mutagenic carcinogens	carcinogens function as aneuploidogens
4 High rates of karyotype-phenotype variations and the origin of ‘immortality’	aneuploidy catalyses karyotype-phenotype variations, including resistance to otherwise lethal conditions, at high rates
5 Cancer-specific aneuploidies	cancer-specific aneuploidies generate cancer phenotypes
6 Complex phenotypes	cancer-specific aneuploidies alter dosages and functions of thousands of genes
7 Nonselective phenotypes	nonselective genes hitchhiking with selective, cancer-specific aneusomies
8 No carcinogenic genes in cancer	cancer is caused by specific aneuploidies

in terms of conventional genetic theories. In the following we offer further commentary on items 1, 2, 5, 6 and 7 listed in table 1, because they are not sufficiently explained by the table and the preceding arguments.

Cancer Is Not Heritable

The chromosomal theory predicts no cancer in newborns, because aneuploidy is not heritable. Aneuploidies are not heritable, because they corrupt embryogenic developmental programs [113, 114], which is usually fatal [157, 191] as originally shown by Boveri [142]. Only some very minor congenital aneuploidies, such as Down syndrome and syndromes based on abnormal numbers of sex chromosomes, are sometimes viable, but only at the cost of severe physiological abnormalities and of no or very low fertility [31, 65, 68, 192]. Thus, ontogenesis is nature’s checkpoint for normal karyotypes. The postnatal exponential increase of the cancer risk with age would then reflect the gradual accumulation of non- or preneoplastic aneuploidy with age, multiplied by the relatively slow, nonselective replication of aneuploid, preneoplastic cells (figs 1, 2).

However, it is as yet unclear, why after initiating doses of carcinogens the neoplastic latencies are very species-dependent, namely much shorter in rodents than in humans [1, 46, 47, 193–195]. It is also unclear, why the increase of the cancer risk is proportional to the lifespan of an animal, i.e. is very low for decades in humans (fig. 1), but only for months in rodents [38, 47]. Still, this is unlikely to be due to species-specific mutation rates, because the rates of conventional mutations are highly conserved in all species [52, 68]. However, the

significantly higher chromosomal instability of aneuploid rodent cells compared to equally aneuploid human cells, shown here in figure 2, may offer a different explanation, namely that chromosomal stability of normal and cancer cells is different in different species.

Long Neoplastic Latencies

The chromosomal evolution theory predicts that carcinogenesis is initially very slow, because preneoplastic cells have no growth advantages compared to normal cells and are typically only little aneuploid (fig. 4). Therefore, they would not form large clonal populations that would increase the probability of further evolutions. The non-clonality of the pre-neoplastic aneuploidies also hides any abnormal phenotypes of pre-neoplastic cells, because phenotypes of single cells are hard to recognize. By contrast, neoplastic ‘progression’ of established cancer cells is predicted to be faster than during the pre-neoplastic phase for two reasons: (1) Neoplastic cells, through their selective phenotypes, will generate large ‘clonal’ populations with high probabilities of further variations. (2) The generally high degrees of most cancer-specific aneuploidies catalyze high rates of chromosomal variations, compared to those of preneoplastic cells (fig. 4).

The chromosomal theory also predicts a certain endpoint of chromosomal evolutions in carcinogenesis. This endpoint would be an equilibrium of aneuploidizations, which is reached once a cancer has maximized cellular variability and adaptability [73] and ‘optimized its genome’ for essential metabolic functions [196]. According to the chromosomal theory maximal chromosomal variability would correspond to near or above triploid chromosome numbers ($>3n$) [13, 73, 137]. Near triploid aneuploidy offers an optimal average redundancy of one spare for each normal chromosome pair, and thus sufficient redundancy to compensate for any losses or genetic mutations of a given chromosome [73]. Accordingly, it is the karyotype of most malignant cancer cells [10, 62, 65, 73, 146, 158, 178, 197].

High Rates of Karyotype-Phenotype Variations and the Origin of Immortality

The chromosomal theory attributes the high rates of karyotype-phenotype variations of cancer cells to the inherent variability of aneuploidy. On this basis, the chromosomal theory also explains the notorious immortality of cancer cells as already described in 1972 by the cytogeneticist Koller [62]: ‘It seems that malignant growth is composed of competing clones of cells with different and continuously changing genotypes, conferring the tumor with an adaptable plasticity against the environment. The bewildering karyotypic patterns reveal the multipotentiality of the neoplastic cell; while normal cells and tissues age and die, through their inherent variability, tumor cells proliferate and survive.’ Thus, cancers are immortal, because subspecies form within the zoos of their polyphyletic

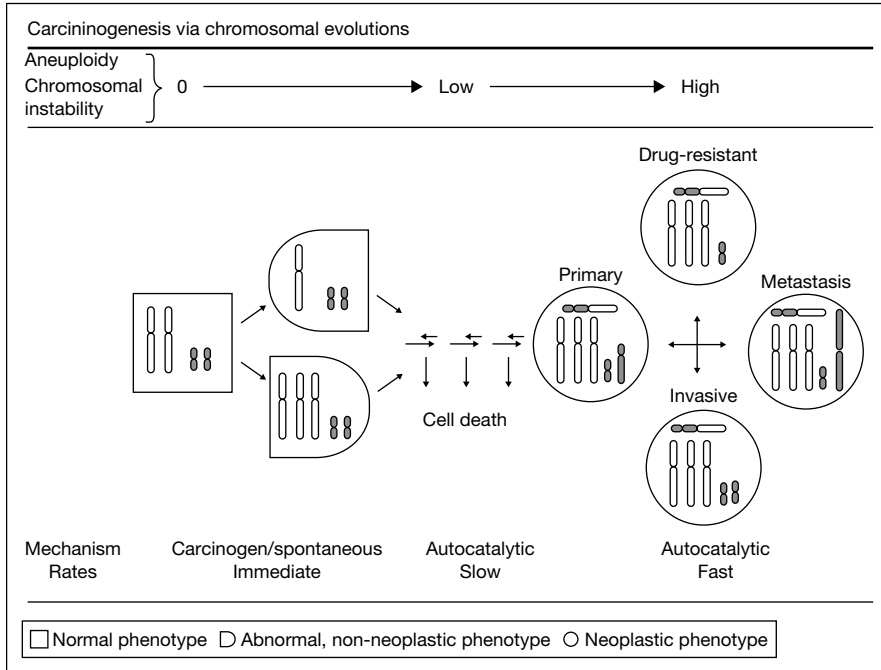


Fig. 4. Carcinogenesis via chromosomal evolutions. According to this mechanism carcinogenesis is initiated by unspecific aneuploidies induced either by carcinogens or spontaneously. Aneuploidy then alters the karyotype automatically at rates that are proportional to the degree of aneuploidy, because it corrupts teams of proteins that segregate, synthesize and repair chromosomes. Aneuploidy is thus a steady source of chromosomal variations from which, in classical Darwinian terms, selection would encourage the evolution and subsequent progressions of neoplastic cell ‘species’ with cancer-specific aneusomies. This evolution would be slow in the preneoplastic phase, because preneoplastic cells have no growth advantages over normal cells and because the degree of preneoplastic aneuploidy is typically low. By comparison the rate of karyotype variations of most cancer cells would be fast, because cancer cells form large populations by outgrowing normal cells and because the degrees of cancer-specific aneuploidy are typically high. Any kind of cancer could have as many specific aneusomies as there are chromosomes involved in the differentiation of its precursor cell in addition to random aneusomies. Thus cancer-specific phenotypes, such as invasiveness, metastasis, and drug-resistance, are generated by the abnormal dosages of thousands of normal genes. Since aneuploidy is inherently unstable, cancer-specific phenotypes, such as drug-resistance, can be reversible or convertible to other specific phenotypes at the same rates at which they are generated. The chromosomal model predicts the heterogeneous phenotypes and karyotypes of cancers as consequences of independent evolutions of the inherently unstable cancer cells. Since aneuploidy causes dedifferentiation, the model further predicts that the degrees of malignancy of cancer cells are proportional to the degrees of aneuploidy.

cell populations [110] – species are defined by karyotypes – survive conditions that are lethal to the mortal majority of the cells, as for example toxic drugs.

Cancer-Specific Aneuploidies

The presence of cancer-specific or nonrandom aneuploidies is directly predicted by and thus correlative proof for the chromosomal theory in terms of Koch's first postulate. Functional proof that cancer-specific aneuploidy generates malignancy could be derived from evidence that the degree of malignancy is proportional to the degree of aneuploidy. Indeed, numerous correlations have confirmed the principle that the degree of malignancy of cancer cells is proportional to their degree of aneuploidy since the 1930s [10, 45, 62–64, 97, 198–204]. Moreover, other studies have shown that maximal malignancy is, indeed, achieved at maximally stable, near-triploid or hypertriploid aneuploidy [65, 178, 197, 205, 206]. The parallel evolutions of aneuploidy and malignancy in cancer cells are thus functional proof for the chromosomal evolution theory of cancer in terms of Koch's third postulate.

Complex Phenotypes

Conventional genetic theories cannot explain the generation of the polygenic cancer-specific phenotypes such as multidrug resistance, polymorphism, metastasis to non-native sites, and transplantability to heterologous species [108] based on conventional rates of mutation and selection in the lifespan of a human or animal. By contrast, the chromosomal theory of cancer explains the complexity of cancer-specific phenotypes by the complexity of the genetic units that are varied, namely chromosomes with thousands of genes. Accordingly, the complex phenotypes of cancer cells have recently been shown to correlate with over- and underexpressions of thousands of genes [34, 87, 116–118, 136]. Likewise, cancer cells over- and underproduce thousands of normal proteins [16, 40, 51, 119].

Nonselective Phenotypes`

Conventional genetic theories explain the evolution of cancer cells by cancer-specific mutations and Darwinian selections. But this mechanism cannot explain the nonselective phenotypes of cancer cells, such as metastasis, drug resistance and 'immortality'. By contrast, the chromosomal theory of carcinogenesis attributes nonselective phenotypes such as metastasis and intrinsic multidrug resistance to nonselective genes hitchhiking with selective, cancer-causing aneusomies, because they are all located on the same chromosomes. The same would be true for that part of acquired multidrug-resistance, which is not directed against the selective drug that induced it. The nonselective phenotype immortality has been explained above.

Conclusions

We conclude that the chromosomal theory provides a coherent explanation of carcinogenesis and can resolve all features of carcinogenesis that are paradoxical in terms of the prevailing genetic theories of cancer. In addition, the theory stands out for making new, clinically testable predictions, as for example the prediction that cancer could be detected prior to malignancy via pre-neoplastic aneuploidy and that chemotherapy could be based on the presence or absence of resistance-specific aneusomies. Thus, if confirmed, the chromosomal theory should become beneficial for cancer research and therapy.

Appendix

The Achilles Heels of the Mutation-Cancer Theory

The currently prevailing cancer theory postulates that cancer is caused by clonal expansion of one single cell that has accumulated about four to seven complementary mutations during the lifetime of a patient [1, 12, 34, 38, 41, 42]. However, the mutation theory is hard to reconcile with the following list of facts.

- 1 *Nonmutagenic Carcinogens.* Contrary to the mutation hypothesis, many carcinogens are not mutagens, including some of the most potent ones. Examples are asbestos, tar, mineral oils, naphthalene, polycyclic aromatic hydrocarbons, butter yellow, urethane, dioxin, hormones, metal ions such as Ni, Cd, Cr, As, spindle blockers such as vincristine and colcemid, extranuclear radiation and solid plastic or metal implants [40, 44, 67, 70, 73, 158, 166, 168].
- 2 *No Transforming Genes.* Despite years of efforts no genes or combinations of genes from cancers have been shown to transform normal cells to cancer cells [4, 5, 138] or mice carrying such genes in their germ lines into polyclonal tumors [1, 24, 56]. Accordingly, many, presumably cancer-specific mutations are not detectably expressed in cancer cells [8, 116, 136, 137].
- 3 *Dependence of Cancer on Unrealistically High Rates of Mutation.* The mutation hypothesis explains the exponential increase of the cancer risk with age by the low probability of four to seven specific mutations [1, 41, 42]. However, in order to maintain the integrity of the genome, spontaneous mutation rates in all species are naturally restricted to about 10^{-7} per dominant gene and to about 10^{-14} per recessive gene per cell generation [6, 47, 52, 57, 68]. Thus, based on these conserved mutation rates cancer via four to seven mutations would not even exist [10]. For example, based on just 4 specific dominant mutations cancer would occur only once in 10^{12} human lifetimes. This is calculated as follows: Since the spontaneous mutation rate per specific, dominant gene is about 10^{-7} , it takes 10^{28} cells to generate one human cell with 4 specific mutations. The expected cancer rate per human lifetime of 1 in 10^{12} is then obtained by dividing 10^{28} by 10^{16} . 10^{16} is the number of cells that correspond to an average human lifetime [10, 38]. Thus, in order to explain the current cancer risk of Americans and Europeans of about 1 in 3 lifetimes [39] (fig. 1), the mutation hypothesis has to assume mutation rates, which are 10^3 [$(10^3)^4 = 10^{12}$] times higher than in conventional mutation.

- 4 *No Explanation for the Long ‘Neoplastic Latency’ in Carcinogenesis Induced by a Critical Dose of Carcinogen.* The mutation hypothesis has no answer to the question why, after a critical dose of carcinogen, carcinogenesis would only occur after exceedingly long ‘neoplastic latencies’ of years to decades [1].
- 5 *Dependence of Phenotype Alterations in Cancers on Unrealistically High Rates of Mutation.* The mutation hypothesis has to assume mutation rates of up to 10^{-3} per cell generation to explain the frequent, spontaneous variation of phenotypes in highly aneuploid cancer cells. Examples are the ‘high rates’, compared to mutation, at which some cancers generate metastatic cells [59, 60], or generate drug-resistant variants [53, 54, 56, 58]. But the mutation rates of most cancers are not higher than those of normal cells [6, 19, 20, 47, 66, 70–74].
- 6 *Heritable Cancer Genes, but no Heritable Cancer.* The four to seven gene mutation hypothesis predicts that subsets of cancer causing mutations should be heritable. Indeed, proponents of the mutation hypothesis have demonstrated that several of the six mutations thought to cause colon cancer [1] can be introduced into the germ line of mice without breaching the viability of these animals. According to one study, animals with one of these mutations, namely ras, were found ‘without detectable phenotypic abnormalities’ [207]. Another study reports, “surprisingly, homozygosity for the Apc1638T mutation is compatible with postnatal life” [208]. Thus subsets of colon cancer genes are heritable. Therefore, colon cancer should be common in newborns, which are clonal for inherited subsets of these six mutations (like transgenic mice). But there is no colon cancer in newborns [38, 39].

Acknowledgments

We thank Harvey Bialy (Institute of Biotechnology, Autonomous National University of Mexico, Cuernavaca, Mexico), Tom Bethell (Washington, D.C.), Alecia DuCharme (UC Berkeley), Siggie Duesberg, George Miklos (Secure Genetics Pty Limited, Sydney, Australia) and Rainer K. Sachs (Departments of Mathematics and of Physics, UC Berkeley) for critical and constructive reviews of the manuscript. We also thank Rainer K. Sachs for figure 1 and David Rasnick (Pretoria, South Africa) for valuable information. We are especially grateful to Robert Leppo (philanthropist, San Francisco) for support and for the fluorescence microscope used for karyotyping of human cancer cells, and the Abraham J. and Phyllis Katz Foundation (New York), an American foundation that prefers to be anonymous, other private sources, and the Forschungsfonds der Fakultät für klinische Medizin Mannheim for steady support.

References

- 1 Vogelstein B, Kinzler KW: The multistep nature of cancer. *Trends Genet* 1993;9:138–141.
- 2 Fujimura J: *Crafting Science: A Sociohistory of the Quest for the Genetics of Cancer*. Cambridge, Harvard University Press, 1996.
- 3 Hahn WC, Counter CM, Lundberg AS, Beijersbergen RL, Brooks MW, Weinberg RA: Creation of human tumour cells with defined genetic elements. *Nature* 1999;400:464–468.
- 4 Li R, Sonik A, Stindl R, Rasnick D, Duesberg P: Aneuploidy versus gene mutation hypothesis of cancer: recent study claims mutation, but is found to support aneuploidy. *Proc Natl Acad Sci USA* 2000;97:3236–3241.
- 5 Li R, Rasnick D, Duesberg P: Correspondence re: Zimonjic D, et al: Derivation of human tumor cells in vitro without widespread genomic instability. *Cancer Res* 2001;61:8838–8844. *Cancer Res* 2002;62:6345–6348; discussion 6348–6349.
- 6 Marx J: Debate surges over the origins of genomic defects in cancer. *Science* 2002;297:544–546.

- 7 Gibbs WW: Untangling the roots of cancer. *Sci Am* 2003;289:56–65.
- 8 Duesberg PH: Are cancers dependent on oncogenes or on aneuploidy? *Cancer Genet Cytogenet* 2003;143:89–91.
- 9 Thilly WG: Have environmental mutagens caused oncomutations in people? *Nat Genet* 2003;34:255–259.
- 10 Duesberg P, Li R: Multistep carcinogenesis: a chain reaction of aneuploidizations. *Cell Cycle* 2003;2:202–210.
- 11 Steinberg D: Appraising aneuploidy as a cancer cause. *Scientist* 2004;18:26–27.
- 12 Radford IR: Chromosomal rearrangement as the basis for human tumourigenesis. *Int J Radiat Biol* 2004;80:543–557.
- 13 Schneider BL, Kulesz-Martin M: Destructive cycles: the role of genomic instability and adaptation in carcinogenesis. *Carcinogenesis* 2004;25:2033–2044.
- 14 Pihan G, Duxsey SJ: Mutations and aneuploidy: co-conspirators in cancer? *Cancer Cell* 2003;4:89–94.
- 15 Zimonjic D, Brooks MW, Popescu N, Weinberg RA, Hahn WC: Correspondence re: Zimonjic D, et al: Derivation of human tumor cells in vitro without widespread genomic instability. *Cancer Res* 2002;62:6348–6349.
- 16 Miklos GLG: The human cancer genome project – one more misstep in the war on cancer. *Nat Biotechnol* 2005;23:535–537.
- 17 Brinkley BR, Goepfert TM: Supernumerary centrosomes and cancer: Boveri's hypothesis resurrected. *Cell Motil Cytoskeleton* 1998;41:281–288.
- 18 Dey P: Aneuploidy and malignancy: an unsolved equation. *J Clin Pathol* 2004;57:1245–1249.
- 19 Tomlinson IP, Novelli MR, Bodmer WF: The mutation rate and cancer. *Proc Natl Acad Sci USA* 1996;93:14800–14803.
- 20 Sieber OM, Heimann K, Tomlinson IP: Genomic instability – the engine of tumorigenesis? *Nat Rev Cancer* 2003;3:701–708.
- 21 Sen S: Aneuploidy and cancer. *Curr Opin Oncol* 2000;12:82–88.
- 22 Duesberg P, Li R, Rasnick D: Aneuploidy approaching a perfect score in predicting and preventing cancer: highlights from a conference held in Oakland in January 2004. *Cell Cycle* 2004;3:823–828.
- 23 Soto AM, Sonnenschein C: The somatic mutation theory of cancer: growing problems with the paradigm? *BioEssays* 2004;26:1097–1107.
- 24 Harris H: A long view of fashions in cancer research. *BioEssays* 2005;27:833–838.
- 25 Duesberg P: Does aneuploidy or mutation start cancer? *Science* 2005;307:41.
- 26 Michor F, Iwasa Y, Vogelstein B, Lengauer C, Nowak MA: Can chromosomal instability initiate tumorigenesis? *Semin Cancer Biol* 2005;15:43–49.
- 27 Varmus HT: Retroviruses and oncogenes, I – Nobel Lecture, Dec 8, 1989. *Biosci Rep* 1990;10:413–430.
- 28 Peltomaki P, Aaltonen LA, Sistonen P, Pylkkanen L, Mecklin J-P, Jarvinen H, Green JS, Jass JR, Weber JL, Leach FS, Petersen GM, Hamilton SR, de la Chapelle A, Vogelstein B: Genetic mapping of a locus predisposing to human colorectal cancer. *Science* 1993;260:810–812.
- 29 Bishop JM: Cancer: the rise of the genetic paradigm. *Genes Dev* 1995;9:1300–1315.
- 30 Vogelstein B, Kinzler KW: *The Genetic Basis of Human Cancer*: New York, McGraw-Hill Health Professions Division, 1998.
- 31 Griffiths AJF, Miller JH, Suzuki DT, Lewontin RC, Gelbart WM: *Genetic analysis*, ed 7. New York, Freeman, 2000.
- 32 Knudson AG: Two genetic hits (more or less) to cancer. *Nat Rev Cancer* 2001;1:157–162.
- 33 Vogelstein B, Kinzler KW: Cancer genes and the pathways they control. *Nat Med* 2004;10:789–799.
- 34 Lodish H, Berk A, Matsudaira P, Kaiser CA, Krieger M, Scott MP, Zipursky SL, Darnell J: *Molecular Cell Biology*, ed 5. New York, Freeman, 2004.
- 35 Pierce BA: *Genetics, a conceptual approach*, ed 2. New York, Freeman, 2005.
- 36 Nordling CO: A new theory on the cancer-inducing mechanism. *Br J Cancer* 1953;7:68–72.
- 37 Armitage P, Doll R: The age distribution of cancer and a multi-stage theory of carcinogenesis. *Br J Cancer* 1954;8:1–12.
- 38 Cairns J: *Cancer: Science and Society*. San Francisco, Freeman, 1978.
- 39 Jemal A, Murray T, Samuels A, Ghafoor A, Ward E, Thun MJ: Cancer statistics, 2003. *CA Cancer J Clin* 2003;53:5–26.

- 40 Pitot HC: *Fundamentals of Oncology*, ed 4. New York, Marcel Dekker, 2002.
- 41 Renan MJ: How many mutations are required for tumorigenesis? Implications from human cancer data. *Mol Carcinog* 1993;7:139–146.
- 42 Hahn WC, Weinberg RA: Modelling the molecular circuitry of cancer. *Nat Rev Cancer* 2002;2:331–341.
- 43 Vogelstein B, Kinzler KW: *The Genetic Basis of Human Cancer*. New York, McGraw-Hill, 1998.
- 44 Berenblum I, Shubik P: An experimental study of the initiating stage of carcinogenesis, and a re-examination of the somatic cell mutation theory of cancer. *Br J Cancer* 1949;3:109–118.
- 45 Foulds L: *Neoplastic Development*. London, Academic Press, 1975.
- 46 Kuroki T, Huh NH: Why are human cells resistant to malignant cell transformation in vitro? *Jpn J Cancer Res* 1993;84:1091–1100.
- 47 Holliday R: Neoplastic transformation: the contrasting stability of human and mouse cells; in Tooze J (eds): *Genetic Instability in Cancer*. Plainview, Cold Spring Harbor Laboratory Press, 1996, vol 28, pp 103–115.
- 48 Rhim JS, Dritschilo A: Neoplastic transformation in human cell culture; in Dritschilo A (ed): *Mechanism of Carcinogenesis*. Totowa, Humana Press, 1991.
- 49 Boice JD Jr, Monson RR: Breast cancer in women after repeated fluoroscopic examinations of the chest. *J Natl Cancer Inst* 1977;59:823–835.
- 50 Doi M, Yukutake M, Tamura K, Watanabe K, Kondo K, Isobe T, Awaya T, Shigenobu T, Oda Y, Yamakido K, Koyama K, Kohno N: A retrospective cohort study on respiratory tract cancers in the workers of the Japanese army poison-gas-factory operated from 1929 to 1945. *Proc Am Soc Clin Oncol* 2002;21:439a.
- 51 Caspersson T: Chemical variability in tumor cell populations. *Acta Unio Int Contra Cancrum* 1964;20:1275–1279.
- 52 Lewin B: *Genes VI*. Oxford, Oxford University Press, 1997.
- 53 Li R, Hehlmann R, Sachs R, Duesberg P: Aneuploidy, the primary cause of the high rates and wide ranges of drug resistance in cancer cells. *Cancer Genet Cytogenet* 2005;in press.
- 54 Duesberg P, Stindl R, Hehlmann R: Explaining the high mutation rates of cancer cells to drug and multidrug resistance by chromosome reassortments that are catalyzed by aneuploidy. *Proc Natl Acad Sci USA* 2000;97:14295–14300.
- 55 Fabarius A, Hehlmann R, Duesberg PH: Instability of chromosome structure in cancer cells increases exponentially with degrees of aneuploidy. *Cancer Genet Cytogenet* 2003;143:59–72.
- 56 Harris H: *The Cells of the Body: A History of Somatic Cell Genetics*. Plainview, Cold Spring Harbor Lab Press, 1995.
- 57 Tlsty TD: Genomic instability and its role in neoplasia; in Kastan MB (ed): *Genetic Instability and Tumorigenesis*. Berlin, Springer, 1997, vol 221, pp 37–46.
- 58 Duesberg P, Stindl R, Hehlmann R: Origin of multidrug resistance in cells with and without multidrug resistance genes: chromosome reassortments catalyzed by aneuploidy. *Proc Natl Acad Sci USA* 2001;98:11283–11288.
- 59 Harris JF, Chambers AF, Hill RP, Ling V: Metastatic variants are generated spontaneously at a high rate in mouse KHT tumor. *Proc Natl Acad Sci USA* 1982;79:5547–5551.
- 60 Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF: Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci USA* 2003;100:3983–3988.
- 61 Heim S, Mitelman F: *Cancer Cytogenetics*, ed 2. New York, Wiley-Liss, 1995.
- 62 Koller PC: *The Role of Chromosomes in Cancer Biology*. New York, Springer, 1972.
- 63 Hauschka TS: The chromosomes in ontogeny and oncogeny. *Cancer Res* 1961;21:957–981.
- 64 Nowell PC: The clonal evolution of tumor cell populations. *Science* 1976;194:23–28.
- 65 Sandberg AA: *The Chromosomes in Human Cancer and Leukemia*, ed 2. New York, Elsevier, 1990.
- 66 Lengauer C, Kinzler KW, Vogelstein B: Genetic instabilities in human cancers. *Nature* 1998;396:643–649.
- 67 Little JB: Radiation carcinogenesis. *Carcinogenesis* 2000;21:397–404.
- 68 Vogel F, Motulsky AG: *Human Genetics: Problems and Approaches*. Berlin, Springer, 1986.
- 69 Tlsty TD: Normal diploid human and rodent cells lack a detectable frequency of gene amplification. *Proc Natl Acad Sci USA* 1990;87:3132–3136.

- 70 Oshimura M, Barrett JC: Chemically induced aneuploidy in mammalian cells: mechanisms and biological significance in cancer. *Environ Mutagen* 1986;8:129–159.
- 71 Strauss BS: The origin of point mutations in human tumor cells. *Cancer Res* 1992;52:249–253.
- 72 Harris CC: Chemical and physical carcinogenesis: advances and perspective for the 1990s. *Cancer Res* 1991;51(18 suppl):5023s–5044s.
- 73 Duesberg P, Fabarius A, Hehlmann R: Aneuploidy, the primary cause of the multilateral genomic instability of neoplastic and preneoplastic cells. *IUBMB Life* 2004;56:65–81.
- 74 Wang TL, Rago C, Silliman N, Ptak J, Markowitz S, Willson JK, Parmigiani G, Kinzler KW, Vogelstein B, Velculescu VE: Prevalence of somatic alterations in the colorectal cancer cell genome. *Proc Natl Acad Sci USA* 2002;99:3076–3080.
- 75 Tomlinson I, Bodmer W: Selection, the mutation rate and cancer: insuring that the tale does not wag the dog. *Nat Med* 1999;5:11–12.
- 76 Zang KD, Singer H: Chromosomal constitution of meningiomas. *Nature* 1967;216:84–85.
- 77 Zang KD: Cytological and cytogenetical studies on human meningioma. *Cancer Genet Cytogenet* 1982;6:249–274.
- 78 Atkin NB, Baker MC: Chromosome abnormalities as primary events in human malignant disease: evidence from marker chromosomes. *J Natl Cancer Inst* 1966;36:539–557.
- 79 Yamamoto T, Rabinowitz Z, Sachs L: Identification of the chromosomes that control malignancy. *Nat New Biol* 1973;243:247–250.
- 80 Oshimura M, Hesterberg TW, Barrett JC: An early, nonrandom karyotypic change in immortal Syrian hamster cell lines transformed by asbestos: trisomy of chromosome 11. *Cancer Genet Cytogenet* 1986;22:225–237.
- 81 Balaban GB, Herlyn M, Clark WH Jr, Nowell PC: Karyotypic evolution in human malignant melanoma. *Cancer Genet Cytogenet* 1986;19:113–122.
- 82 Atkin NB: Chromosome 1 aberrations in cancer. *Cancer Genet Cytogenet* 1986;21:279–285.
- 83 Atkin NB: Non-random chromosomal changes in human neoplasia; in Obe G (ed): *Eukaryotic Chromosomes: Structural and Functional Aspects*. New Delhi, Narosa, 1991, pp 153–164.
- 84 Fabarius A, Willer A, Yerganian G, Hehlmann R, Duesberg P: Specific aneusomies in Chinese hamster cells at different stages of neoplastic transformation, initiated by nitrosomethylurea. *Proc Natl Acad Sci USA* 2002;99:6778–6783.
- 85 Pejonic T, Heim S, Oerndal C, Jin Y, Mandahl N, Willen H, Mitelman F: Simple numerical chromosome aberrations in well-defined malignant epithelial tumors. *Cancer Genet Cytogenet* 1990;49:95–101.
- 86 Johansson B, Bardi G, Pandis N, Gorunova L, Backman PL, Mandahl N, Dawiskiba S, Andren-Sandberg A, Heim S, Mitelman F: Karyotypic pattern of pancreatic adenomas correlates with survival and tumor grade. *Int J Cancer* 1994;58:8–13.
- 87 Virtaneva K, Wright FA, Tanner SM, Yuan B, Lemon WJ, Caligiuri MA, Bloomfield CD, de La Chapelle A, Krahe R: Expression profiling reveals fundamental biological differences in acute myeloid leukemia with isolated trisomy 8 and normal cytogenetics. *Proc Natl Acad Sci USA* 2001;98:1124–1129.
- 88 Petersen I, Langreck H, Wolf G, Schwendel A, Psille R, Vogt P, Reichel MB, Ried T, Dietel M: Small-cell lung cancer is characterized by a high incidence of deletions on chromosomes 3p, 4q, 5q, 10q, 13q and 17p. *Br J Cancer* 1997;75:79–86.
- 89 Ried T, Heselmeyer-Haddad K, Blegen H, Schrock E, Auer G: Genomic changes defining the genesis, progression, and malignancy potential in solid human tumors: a phenotype/genotype correlation. *Genes Chromosomes Cancer* 1999;25:195–204.
- 90 Jiang F, Richter J, Schraml P, Bubendorf L, Gasser T, Sauter G, Mihatsch MJ, Moch H: Chromosomal imbalances in papillary renal cell carcinoma: genetic differences between histological subtypes. *Am J Pathol* 1998;153:1467–1473.
- 91 Richter J, Beffa L, Wagner U, Schraml P, Gasser TC, Moch H, Mihatsch MJ, Sauter G: Patterns of chromosomal imbalances in advanced urinary bladder cancer detected by comparative genomic hybridization. *Am J Pathol* 1998;153:1615–1621.
- 92 Nupponen NN, Kakkola L, Koivisto P, Visakorpi T: Genetic alterations in hormone-refractory recurrent prostate carcinomas. *Am J Pathol* 1998;153:141–148.

- 93 Weber RG, Scheer M, Born IA, Joos S, Cobbers JM, Hofele C, Reifenberger G, Zoller JE, Lichter P: Recurrent chromosomal imbalances detected in biopsy material from oral premalignant and malignant lesions by combined tissue microdissection, universal DNA amplification, and comparative genomic hybridization. *Am J Pathol* 1998;153:295–303.
- 94 Gebhart E, Liehr T: Patterns of genomic imbalances in human solid tumors. *Int J Oncol* 2000;16:383–399.
- 95 Dellas A, Torhorst J, Jiang F, Proffitt J, Schultheiss E, Holzgreve W, Sauter G, Mihatsch MJ, Moch H: Prognostic value of genomic alterations in invasive cervical squamous cell carcinoma of clinical stage IB detected by comparative genomic hybridization. *Cancer Res* 1999;59:3475–3479.
- 96 Patel AS, Hawkins AL, Griffin CA: Cytogenetics and cancer. *Curr Opin Oncol* 2000;12:62–67.
- 97 Wilkens L, Flemming P, Gebel M, Bleck J, Terkamp C, Wingen L, Kreipe H, Schlegelberger B: Induction of aneuploidy by increasing chromosomal instability during dedifferentiation of hepatocellular carcinoma. *Proc Natl Acad Sci USA* 2004;101:1309–1314.
- 98 Hoglund M, Sall T, Heim S, Mitelman F, Mandahl N, Fadl-Elmula I: Identification of cytogenetic subgroups and karyotypic pathways in transitional cell carcinoma. *Cancer Res* 2001;61:8241–8246.
- 99 Heselmeyer-Haddad K, Sommerfeld K, White NM, Chaudhri N, Morrison LE, Palanisamy N, Wang ZY, Auer G, Steinberg W, Ried T: Genomic amplification of the human telomerase gene (TERC) in pap smears predicts the development of cervical cancer. *Am J Pathol* 2005;166:1229–1238.
- 100 Meijer GA, Hermsen MA, Baak JP, van Diest PJ, Meuwissen SG, Belien JA, Hoovers JM, Joenje H, Snijders PJ, Walboomers JM: Progression from colorectal adenoma to carcinoma is associated with non-random chromosomal gains as detected by comparative genomic hybridisation. *J Clin Pathol* 1998;51:901–909.
- 101 Aragane H, Sakakura C, Nakanishi M, Yasuoka R, Fujita Y, Taniguchi H, Hagiwara A, Yamaguchi T, Abe T, Inazawa J, Yamagishi H: Chromosomal aberrations in colorectal cancers and liver metastases analyzed by comparative genomic hybridization. *Int J Cancer* 2001;94:623–629.
- 102 Nakao K, Shibusawa M, Ishihara A, Yoshizawa H, Tsunoda A, Kusano M, Kurose A, Makita T, Sasaki K: Genetic changes in colorectal carcinoma tumors with liver metastases analyzed by comparative genomic hybridization and DNA ploidy. *Cancer* 2001;91:721–726.
- 103 Al-Mulla F, Keith WN, Pickford IR, Going JJ, Birnie GD: Comparative genomic hybridization analysis of primary colorectal carcinomas and their synchronous metastases. *Genes Chromosomes Cancer* 1999;24:306–314.
- 104 Hermsen M, Postma C, Baak J, Weiss M, Rapallo A, Sciuotto A, Roemen G, Arends JW, Williams R, Giaretti W, De Goeij A, Meijer G: Colorectal adenoma to carcinoma progression follows multiple pathways of chromosomal instability. *Gastroenterology* 2002;123:1109–1119.
- 105 Knosel T, Schluns K, Stein U, Schwabe H, Schlag PM, Dietel M, Petersen I: Chromosomal alterations during lymphatic and liver metastasis formation of colorectal cancer. *Neoplasia* 2004;6:23–28.
- 106 Petersen I, Petersen S: Towards a genetic-based classification of human lung cancer. *Anal Cell Pathol* 2001;22:111–121.
- 107 Schimke RT: Gene amplification, drug resistance, and cancer. *Cancer Res* 1984;44:1735–1742.
- 108 Hauschka TS, Levan A: Inverse relationship between chromosome ploidy and host-specificity of sixteen transplantable tumors. *Exp Cell Res* 1953;4:457–467.
- 109 Vogt M: A study of the relationship between karyotype and phenotype in cloned lines of strain HeLa. *Genetics* 1959;44:1257–1270.
- 110 Hauschka T, Levan A: Cytologic and functional characterization of single cell clones isolated from the Krebs-2 and Ehrlich ascites tumors. *J Natl Cancer Inst* 1958;21:77–111.
- 111 Reeves RH: Recounting a genetic story. *Nature* 2000;405:283–284.
- 112 Mao R, Zielke CL, Zielke HR, Pevsner J: Global up-regulation of chromosome 21 gene expression in the developing Down syndrome brain. *Genomics* 2003;81:457–467.
- 113 Shapiro BL: Down syndrome: a disruption of homeostasis. *Am J Med Genet* 1983;14:241–269.
- 114 Epstein C: *The Consequences of Chromosome Imbalance: Principles, Mechanisms, and Models*. Cambridge, Cambridge University Press, 1986.
- 115 Schoenlein PV: Molecular cytogenetics of multiple drug resistance. *Cytotechnology* 1993;12:63–89.
- 116 Pollack JR, Sorlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE, Tibshirani R, Botstein D, Borresen-Dale AL, Brown PO: Microarray analysis reveals a major direct role of DNA copy number

- alteration in the transcriptional program of human breast tumors. *Proc Natl Acad Sci USA* 2002;99:12963–12968.
- 117 Furge KA, Lucas KA, Takahashi M, Sugimura J, Kort EJ, Kanayama HO, Kagawa S, Hoekstra P, Curry J, Yang XJ, Teh BT: Robust classification of renal cell carcinoma based on gene expression data and predicted cytogenetic profiles. *Cancer Res* 2004;64:4117–4121.
 - 118 Aggarwal A, Leong SH, Lee C, Kon OL, Tan P: Wavelet transformations of tumor expression profiles reveals a pervasive genome-wide imprinting of aneuploidy on the cancer transcriptome. *Cancer Res* 2005;65:186–194.
 - 119 Caspersson T, Foley GE, Killander D, Lomakka G: Cytochemical differences between mammalian cell lines of normal and neoplastic origins: correlation with heterotransplantability in Syrian hamsters. *Exp Cell Res* 1963;32:553–565.
 - 120 Pihan GA, Purohit A, Wallace J, Knecht H, Woda B, Quesenberry P, Doxsey SJ: Centrosome defects and genetic instability in malignant tumors. *Cancer Res* 1998;58:3974–3985.
 - 121 Lingle WL, Barrett SL, Negron VC, D'Assoro AB, Boeneman K, Liu W, Whitehead CM, Reynolds C, Salisbury JL: Centrosome amplification drives chromosomal instability in breast tumor development. *Proc Natl Acad Sci USA* 2002;99:1978–1983.
 - 122 Pihan GA, Wallace J, Zhou Y, Doxsey SJ: Centrosome abnormalities and chromosome instability occur together in pre-invasive carcinomas. *Cancer Res* 2003;63:1398–1404.
 - 123 Ghadimi BM, Sackett DL, Difilippantonio MJ, Schrock E, Neumann T, Jauho A, Auer G, Ried T: Centrosome amplification and instability occurs exclusively in aneuploid, but not in diploid colorectal cancer cell lines, and correlates with numerical chromosomal aberrations. *Genes Chromosomes Cancer* 2000;27:183–190.
 - 124 Lingle WL, Lutz WH, Ingle JN, Maihle NJ, Salisbury JL: Centrosome hypertrophy in human breast tumors: implications for genomic stability and cell polarity. *Proc Natl Acad Sci USA* 1998;95:2950–2955.
 - 125 Doxsey S, Zimmerman W, Mikule K: Centrosome control of the cell cycle. *Trends Cell Biol* 2005;15:303–311.
 - 126 Bernards R, Weinberg RA: A progression puzzle. *Nature* 2002;418:823.
 - 127 Goldie JH: Drug resistance in cancer: a perspective. *Cancer Metastasis Rev* 2001;20:63–68.
 - 128 Doubre H, Cesari D, Mairovitz A, Benac C, Chantot-Bastarud S, Dagnon K, Antoine M, Danel C, Bernaudin JF, Fleury-Feith J: Multidrug resistance-associated protein (MRP1) is overexpressed in DNA aneuploid carcinomatous cells in non-small cell lung cancer (NSCLC). *Int J Cancer* 2005;113:568–574.
 - 129 Hayflick L, Moorhead PS: The serial cultivation of human diploid cell strains. *Exp Cell Res* 1961;25:585–621.
 - 130 Bishop JM: Enemies within: genesis of retrovirus oncogenes. *Cell* 1981;23:5–6.
 - 131 Tabin CJ, Bradley SM, Bargmann CI, Weinberg RA, Papageorge AG, Scolnick EM, Dhar R, Lowy DR, Chang EH: Mechanism of activation of a human oncogene. *Nature* 1982;300:143–149.
 - 132 Bishop JM: The molecular genetics of cancer. *Science* 1987;235:305–311.
 - 133 Varmus HE: The molecular genetics of cellular oncogenes. *Annu Rev Genet* 1984;18:533–612.
 - 134 Boland CR, Ricciardello L: How many mutations does it take to make a tumor? *Proc Natl Acad Sci USA* 1999;96:14675–14677.
 - 135 Kallioniemi OP, Kallioniemi A, Kurisu W, Thor A, Chen LC, Smith HS, Waldman FM, Pinkel D, Gray JW: ERBB2 amplification in breast cancer analyzed by fluorescence in situ hybridization. *Proc Natl Acad Sci USA* 1992;89:5321–5325.
 - 136 Zhang L, Zhou W, Velculescu VE, Kern SE, Hruban RH, Hamilton SR, Vogelstein B, Kinzler KW: Gene expression profiles in normal and cancer cells. *Science* 1997;276:1268–1272.
 - 137 Rasnick D, Duesberg P: How aneuploidy affects metabolic control and causes cancer. *Biochem J* 1999;340:621–630.
 - 138 Akagi T, Sasai K, Hanafusa H: Refractory nature of normal human diploid fibroblasts with respect to oncogene-mediated transformation. *Proc Natl Acad Sci USA* 2003;100:13567–13572.
 - 139 Atkin NB, Baker MC: Are human cancers ever diploid – or often trisomic? Conflicting evidence from direct preparations and cultures. *Cytogenet Cell Genet* 1990;53:58–60.
 - 140 Duesberg P, Rausch C, Rasnick D, Hehlmann R: Genetic instability of cancer cells is proportional to their degree of aneuploidy. *Proc Natl Acad Sci USA* 1998;95:13692–13697.

- 141 Hanseemann D: Ueber asymmetrische Zelltheilung in Epithelkrebsen und deren biologische Bedeutung. *Virchows Arch Pathol Anat* 1890;119:299–326.
- 142 Boveri T: On multipolar mitosis as a means of analysis of the cell nucleus; in Willier BH, Oppenheimer JM (eds): *Foundations of Experimental Embryology*. New York, Prentice Hall, 1902/1964, pp 74–97.
- 143 Boveri T: *Zur Frage der Entstehung maligner Tumoren*. Jena, Fischer, 1914.
- 144 Darlington CD: Plasmogene theory and cancer genesis; in *Genetics and Cancer; A collection of papers presented at the thirteenth annual symposium on fundamental cancer research, 1959*. Austin, University of Texas MD Anderson Hospital and Tumor Institute, 1959, pp 9–24.
- 145 Rous P: Surmise and fact on the nature of cancer. *Nature* 1959;183:1357–1361.
- 146 Johansson B, Mertens F, Mitelman F: Primary vs. secondary neoplasia-associated chromosomal abnormalities – balanced rearrangements vs. genomic imbalances? *Genes Chromosomes Cancer* 1996;16:155–163.
- 147 Levan A, Levan G, Mitelman F: Chromosomes and cancer. *Hereditas* 1977;86:15–30.
- 148 Pennisi E: Trigger for centrosome replication found. *Science* 1999;238:770–771.
- 149 Grimm D: Genetics. Disease backs cancer origin theory. *Science* 2004;306:389.
- 150 Lengauer C, Wang Z: From spindle checkpoint to cancer. *Nat Genet* 2004;36:1144–1145.
- 151 Rajagopalan H, Lengauer C: Aneuploidy and cancer. *Nature* 2004;432:338–341.
- 152 Hede K: Which came first? Studies clarify role of aneuploidy in cancer. *J Natl Cancer Inst* 2005;97:87–89.
- 153 Rajagopalan H, Jallepalli PV, Rago C, Velculescu VE, Kinzler KW, Vogelstein B, Lengauer C: Inactivation of hCDC4 can cause chromosomal instability. *Nature* 2004;428:77–81.
- 154 O'Brien S, Menotti-Raymond M, Murphy W, Nash W, Wornberg J, Stanyon R, Copeland N, Jenkins N, Womack J, Marshall Graves J: The promise of comparative genomics in mammals. *Science* 1999;286:458–481.
- 155 Lindsley DL, Sandler L, Baker BS, Carpenter ATC, Denell RE, Hall JC, Jacobs PA, Gabor Miklos GL, Davis BK, Gethmann RC, Hardy RW, Hessler A, Miller SM, Nozawa H, Parry DM, Gould-Somero M: Segmental aneuploidy and the genetic gross structure of the *Drosophila* genome. *Genetics* 1972;71:157–184.
- 156 Matzke MA, Mittelsten-Scheid O, Matzke AJM: Rapid structural and epigenetic changes in polyploid and aneuploid genomes. *BioEssays* 1999;21:761–767.
- 157 Hernandez D, Fisher EM: Mouse autosomal trisomy: two's company, three's a crowd. *Trends Genet* 1999;15:241–247.
- 158 Duesberg P, Rasnick D: Aneuploidy, the somatic mutation that makes cancer a species of its own. *Cell Motil Cytoskeleton* 2000;47:81–107.
- 159 Sachidanandam R, Weissman D, et al: A map of human genome sequence variation containing 1.42 million single nucleotide polymorphisms. *Nature* 2001;409:928–933.
- 160 Kinzler KW, Vogelstein B: Gatekeepers and caretakers. *Nature* 1997;386:761–762.
- 161 Kacser H, Burns JA: Molecular democracy: who shares the controls? *Biochem Soc Trans* 1979;7:1149–1160.
- 162 Hartman JL, Garvik B, Hartwell L: Principles for the buffering of genetic variation. *Science* 2001;291:1001–1004.
- 163 Cornish-Bowden A: Metabolic control analysis in biotechnology and medicine. *Nature Biotechnology* 1999;17:641–643.
- 164 Ames B, Durston WE, Yamaski E, Lee FD: Carcinogens are mutagens: a simple test system combining liver homogenates for activation and bacteria for detection. *Proc Natl Acad Sci USA* 1973;70:2281–2285.
- 165 Alberts B, Bray D, Lewis J, Raff M, Roberts K, Watson JD: *Molecular Biology of the Cell*. New York, Garland, 1994.
- 166 Burdette WJ: The significance of mutation in relation to origin of tumors. *Cancer Res* 1955;15:201–226.
- 167 Conti JC, Aldaz CM, O'Connell J, Klen-Szanto AJ-P, Slaga TJ: Aneuploidy, an early event in mouse skin tumor development. *Carcinogenesis* 1986;7:1845–1848.
- 168 Lijinsky W: A view of the relation between carcinogenesis and mutagenesis. *Environ Mol Mutagen* 1989;14(suppl 16):78–84.

- 169 Li R, Yerganian G, Duesberg P, Kraemer A, Willer A, Rausch C, Hehlmann R: Aneuploidy correlated 100% with chemical transformation of Chinese hamster cells. *Proc Natl Acad Sci USA* 1997;94:14506–14511.
- 170 Duesberg P, Li R, Rasnick D, Rausch C, Willer A, Kraemer A, Yerganian G, Hehlmann R: Aneuploidy precedes and segregates with chemical carcinogenesis. *Cancer Genet Cytogenet* 2000;119:83–93.
- 171 Kadhim MA, Macdonald DA, Goodhead DT, Lorimore SA, Marsden SJ, Wright EG: Transmission of chromosomal instability after plutonium α -particle irradiation. *Nature* 1992;355:738–740.
- 172 Holmberg K, Falt S, Johansson A, Lambert B: Clonal chromosome aberrations and genomic instability in X-irradiated human T-lymphocyte cultures. *Mutat Res* 1993;286:321–330.
- 173 Trott DA, Cuthbert AP, Overell RW, Russo I, Newbold RF: Mechanisms involved in the immortalization of mammalian cells by ionizing radiation and chemical carcinogens. *Carcinogenesis* 1995;16:193–204.
- 174 Wright EG: Inherited and inducible chromosomal instability: a fragile bridge between genome integrity mechanisms and tumorigenesis. *J Pathol* 1999;187:19–27.
- 175 Roschke AV, Tonon G, Gehlhaus KS, McTyre N, Bussey KJ, Lababidi S, Scudiero DA, Weinstein JN, Kirsch IR: Karyotypic complexity of the NCI-60 drug-screening panel. *Cancer Res* 2003;63:8634–8647.
- 176 Kost-Alimova M, Fedorova L, Yang Y, Klein G, Imreh S: Microcell-mediated chromosome transfer provides evidence that polysomy promotes structural instability in tumor cell chromosomes through asynchronous replication and breakage within late-replicating regions. *Genes Chromosomes Cancer* 2004;40:316–324.
- 177 Camps J, Ponsa I, Ribas M, Prat E, Egozcue J, Peinado MA, Miro R: Comprehensive measurement of chromosomal instability in cancer cells: combination of fluorescence in situ hybridization and cytokinesis-block micronucleus assay. *FASEB J* 2005;19:828–830.
- 178 Lengauer C, Kinzler KW, Vogelstein B: Genetic instability in colorectal cancers. *Nature* 1997;386:623–627.
- 179 Das SK, Kunkel TA, Loeb LA: Effects of altered nucleotide concentrations on the fidelity of DNA replication. *Basic Life Sci* 1985;31:117–126.
- 180 Knudson AG: Chasing the cancer demon. *Annu Rev Genet* 2000;34:1–19.
- 181 Hoeijmakers JH: Genome maintenance mechanisms for preventing cancer. *Nature* 2001;411:366–374.
- 182 German J: Bloom's syndrome. II. The prototype of genetic disorders predisposing to chromosome instability and cancer; in German J (ed): *Chromosomes and Cancer*. New York, Wiley, 1974, pp 601–617.
- 183 Hanks S, Coleman K, Reid S, Plaja A, Firth H, Fitzpatrick D, Kidd A, Mehes K, Nash R, Robin N, Shannon N, Tolmie J, Swansbury J, Irrthum A, Douglas J, Rahman N: Constitutional aneuploidy and cancer predisposition caused by biallelic mutations in BUB1B. *Nat Genet* 2004;36:1159–1161.
- 184 Kajii T, Kawai T, Takumi T, Misu H, Mabuchi O, Takahashi Y, Tachino M, Nihei F, Ikeuchi T: Mosaic variegated aneuploidy with multiple congenital abnormalities: homozygosity for total premature chromatid separation trait. *Am J Med Genet* 1998;78:245–249.
- 185 Lanza A, Lagomarsini P, Casati A, Ghetti P, Stefanini M: Chromosomal fragility in the cancer-prone disease xeroderma pigmentosum preferentially involves bands relevant for cutaneous carcinogenesis. *Int J Cancer* 1997;74:654–663.
- 186 Benedict WF, Banerjee A, Mark C, Murphree AL: Non-random retinoblastoma gene is a recessive cancer gene. *Cancer Genet Cytogenet* 1983;10:311–333.
- 187 Gardener HA, Gallie BL, Knight LA, Phillips RA: Multiple karyotypic changes in retinoblastoma tumor cells: presence of normal chromosome No. 13 in most tumors. *Cancer Genet Cytogenet* 1982;6:201–211.
- 188 Hamel PA, Phillips RA, Muncaster M, Gallie BL: Speculations on the roles of RB1 in tissue-specific differentiation, tumor initiation, and tumor progression. *FASEB J* 1993;7:846–854.
- 189 Amare Kadam PS, Ghule P, Jose J, Bamne M, Kurkure P, Banavali S, Sarin R, Advani S: Constitutional genomic instability, chromosome aberrations in tumor cells and retinoblastoma. *Cancer Genet Cytogenet* 2004;150:33–43.

- 190 Worsham MJ, Carey TE, Benninger MS, Gasser KM, Kelker W, Zarbo RJ, Van Dyke DL: Clonal cytogenetic evolution in a squamous cell carcinoma of the skin from a xeroderma pigmentosum patient. *Genes Chromosomes Cancer* 1993;7:158–164.
- 191 Hassold TJ: Chromosome abnormalities in human reproductive wastage. *Trends Genet* 1986;2:105–110.
- 192 Bauer KH: *Das Krebsproblem*, ed 2. Berlin, Springer, 1963.
- 193 Fusenig NE, Boukamp P: Multiple stages and genetic alterations in immortalization, malignant transformation, and tumor progression of human skin keratinocytes. *Mol Carcinog* 1998;23:144–158.
- 194 Urano K, Katakai Y, Tokuda Y, Ueyama Y, Nomura T, Yamamoto S: Failure of genotoxic carcinogens to produce tumors in human skin xenografts transplanted to SCID mice. *Carcinogenesis* 1995;16:2223–2226.
- 195 Soballe PW, Montone KT, Satyamoorthy K, Nesbit M, Herlyn M: Carcinogenesis in human skin grafted to SCID mice. *Cancer Res* 1996;56:757–764.
- 196 Roschke AV, Stover K, Tonon G, Schaffer AA, Kirsch IR: Stable karyotypes in epithelial cancer cell lines despite high rates of ongoing structural and numerical chromosomal instability. *Neoplasia* 2002;4:19–31.
- 197 Atkin NB: Nuclear size in premalignant conditions of the cervix uteri. *Nature* 1964;202:201.
- 198 Winge O: Zytologische Untersuchungen ueber die Natur maligner Tumoren. II. Teerkarzinome bei Maesen. *Z Zellforsch Mikrosk Anat* 1930;10:683–735.
- 199 Spriggs AI: Cytogenetics of cancer and precancerous states of the cervix uteri; in German J (ed): *Chromosomes and Cancer*. New York, Wiley, 1974, p 423.
- 200 Frankfurt OS, Chin JL, Englander LS, Greco WR, Pontes JE, Rustum YM: Relationship between DNA ploidy, glandular differentiation, and tumor spread in human prostate cancer. *Cancer Res* 1985;45:1418–1423.
- 201 Bocking A, Chatelain R: Diagnostic and prognostic value of DNA cytometry in gynecologic cytology. *Anal Quant Cytol Histol* 1989;11:177–186.
- 202 Choma D, Daures JP, Quantin X, Pujol JL: Aneuploidy and prognosis of non-small-cell lung cancer: a meta-analysis of published data. *Br J Cancer* 2001;85:14–22.
- 203 Schoch C, Haferlach T, Haase D, Fonatsch C, Loffler H, Schlegelberger B, Staib P, Sauerland MC, Heinecke A, Buchner T, Hiddemann W: Patients with de novo acute myeloid leukaemia and complex karyotype aberrations show a poor prognosis despite intensive treatment: a study of 90 patients. *Br J Haematol* 2001;112:118–126.
- 204 Grimwade D, Walker H, Harrison G, Oliver F, Chatters S, Harrison CJ, Wheatley K, Burnett AK, Goldstone AH: The predictive value of hierarchical cytogenetic classification in older adults with acute myeloid leukemia (AML): analysis of 1065 patients entered into the United Kingdom Medical Research Council AML11 trial. *Blood* 2001;98:1312–1320.
- 205 Giaretti W: A model of DNA aneuploidization and evolution in colorectal cancer. *Lab Invest* 1994;71:904–910.
- 206 Shackney SE, Berg G, Simon SR, Cohen J, Amina S, Pommersheim W, Yakulis R, Wang S, Uhl M, Smith CA, Pollice A, Hartsock R: Origins and clinical implications of aneuploidy in early bladder cancer. *Cytometry* 1995;22:307–316.
- 207 Kim SH, Roth KA, Moser AR, Gordon JI: Transgenic mouse models that explore the multistep hypothesis of intestinal neoplasia. *J Cell Biol* 1993;123:877–893.
- 208 Smits R, Kielman MF, Breukel C, Zurcher C, Neufeld K, Jagmohan-Changur S, Hofland N, van Dijk J, White R, Edelmann W, et al.: Apc1638T: a mouse model delineating critical domains of the adenomatous polyposis coli protein involved in tumorigenesis and development. *Genes Dev* 1999;13:1309–1321.

Peter Duesberg, PhD
 Department of Molecular and Cellular Biology
 Donner Laboratory, UC Berkeley
 Berkeley, CA (USA)
 Tel. +1 510 642 5649, Fax +1 510 642 6455, E-Mail duesberg@berkeley.edu